1	Heritability and genetic architecture of reproduction-related traits in a
2	temperate oak species
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11 Abstract

12 Reproduction, one of the main component of plant fitness, is highly variable in response to 13 environmental cues, but little is known about the genetic determinism underlying reproduction-14 related traits in forest tree species. There is therefore an urgent need to characterize the genetic 15 architecture of those traits if we are to predict the evolutionary trajectories of forest populations facing 16 rapidly changing environment and mitigate their impacts. Using a full-sib family of pedunculate oak 17 (Quercus robur), we investigated the within population variability of seed production and mean seed 18 mass during four consecutive years. Reproductive traits were highly variable between trees and 19 between years. The high narrow sense heritability and evolvability estimated underline the important 20 genetic effect on the variability in seed production and mean seed mass. Despite a large variability 21 over years, reproductive traits show significant genetic correlation between years. Furthermore, for 22 the first time in forest tree species, quantitative trait loci (QTLs) associated with seed production and 23 mean mass of a seed have been identified. While it is commonly assumed and observed that fitness-24 traits have low narrow sense heritabilities, our findings show that reproduction-related traits may 25 undergo evolutionary changes under selective pressure and may be determinant for tree adaptation.

26 <u>Keywords</u>: Tree reproduction, Seed production, Fitness, Heritability, QTLs, *Quercus robur*

27 1. Introduction

28 Tree fitness defines the ability of an individual to survive, grow and reproduce in an environment. 29 Numerous studies have investigated to which extent the phenotypic variation of growth and survival 30 in response to environmental changes was genetically or environmentally driven in forest tree species 31 but there is a lack of studies investigating reproduction (Kang et al., 2003; Santos del Blanco et al., 32 2010; Santos-del-Blanco et al., 2012). Yet, understanding the variability in tree reproduction is essential 33 in order to assess the process of adaptation of forest tree species in response to environmental 34 changes (Aitken et al., 2008; Anderson, 2016). Increased reproduction enhances fitness of trees 35 through different processes. For example, higher seed production increases the probability of 36 dispersion by animals (Howe and Smallwood, 1982; Schupp et al., 2010; Traveset et al., 2014) and thus 37 enhances population growth and dynamics. Long-distance dispersion generally due to scatter-38 hoarding, i.e. the way animals buried their seeds for latter consumption, favors regeneration because 39 (1) seed dispersed escape density-dependent competition under the mother tree and (2) because 40 during years of abundant seed production, buried seeds will not all be consumed by predators (Vander 41 Wall, 2010). Furthermore, the variability in seed size within a same species also plays a role as it is 42 correlated with the germination rate (Gómez, 2004; Walters and Reich, 2000). Thus, because of the 43 larger amount of reserves, bigger seed are more likely to survive to infestation from insects and 44 seedlings' early development is favored (Bonal et al., 2007; Sousa et al., 2003).

45 Tree reproduction is highly variable in response to environmental changes. Previous studies have 46 shown large variations in seed production, seed size or seed germination along environmental 47 gradients and across years (in oaks Caignard et al., 2017; Koenig et al., 2009; in pine Lopez-Toledo et 48 al., 2017; in black spruce Sirois, 2000). Part of this variation is driven by environmental changes such 49 as temperature and precipitation. For example, seed production in European oak species increases 50 with a rise in temperature during the period of pollination (Caignard et al., 2017; Schermer et al. 51 submitted) and decreases with increasing drought occurring during late summer (Bogdziewicz et al., 52 2017; Pérez-Ramos et al., 2010; Sanchez-Humanes and Espelta, 2011). While numerous studies have investigated the phenotypic variability of reproduction traits in response to environmental changes
little is known about the genetic source of this variation.

55 Common garden experiments have been useful in documenting genetically based 56 differentiationamong and within populations but few have focused on the genetic variations of 57 reproductive traits, most likely because their assessments can only be made on reproductively mature 58 trees while other fitness related traits can be measured on juvenile traits. In comparison, the genetics 59 of reproductive traits have been already explored for annual plants (Alonso-Blanco et al., 1999; 60 Mitchell-Olds, 1996), plant crops (Doligez et al., 2002; Houel et al., 2015; Kadri et al., 2017) and fruit trees (Guitton et al., 2011; Sadok et al., 2013; Wu et al., 2014). For example, quantitative traits loci 61 62 (QTLs) associated with seed size, seed number or size at first reproduction were detected for Arabidopsis thaliana (Alonso-Blanco et al., 1999) and Vitis vinifera (Doligez et al., 2002). In fruit trees, 63 64 genetic investigations were oriented towards flowering and fruiting phenology (Dirlewanger et al., 65 2012; Romeu et al., 2014) and biennial fruit bearing, i.e. the yearly alternation of flowering and fruiting 66 (Guitton et al., 2011; Sadok et al., 2013; Shalom et al., 2012). In forest trees, the phenological variation 67 of the apical buds has been extensively investigated in recent years (Derory et al., 2010; Jermstad et al., 2001; Pelgas et al., 2011; Scotti-Saintagne et al., 2004) but very few studies have targeted the 68 69 genetic variation of reproduction within population (Bilir et al., 2006; Sivacioglu et al., 2009; 70 Tsubomura et al., 2012) and even fewer have attempted to detect QTLs underlying reproductive traits 71 (see Ujino-Ihara et al., 2012 for male strobilus abundance). Finally, Pearse et al. (2016) strongly 72 advocated for assessing heritability of masting related traits, while recalling that relevant data may 73 already exist (El Kassaby and Barclay, 1992). As the selective response to environmental changes, and 74 so the capacity of adaptation of tree population largely depend on the genetic variations and the 75 genetic architecture underlying fitness related traits, it is necessary to investigate more reproduction.

In this study we assessed the phenotypic and genetic variation of reproductive traits and searched
for their underlying quantitative trait loci (QTL) in a full-sib family of a broadly distributed temperate
oak species (*Quercus robur L.*). Our main objectives were to (i) subdivide the total phenotypic variation

of reproductive traits into environmental and genetic components (ii) to dissect the genetic architecture of reproductive traits by detecting their underlying QTLs (iii) to examine the stability of QTLs expression over successive years.

82 2. Material and Methods

83 2.1. Experimental design

84 A full-sib family originating from a controlled cross of pedunculate oak (Quercus robur) was 85 used. The male parent (A4) was selected in a park close to Arcachon in the South West of France (44°40'N, 1°11'W) and the female parent (3P) originated from the INRA forest research station of 86 Pierroton (44°44'N, 0°46'W). The controlled cross was carried out in 1992 and provided 278 seedlings, 87 88 which subsequently were vegetatively propagated. Rooted cuttings from 207 genotypes (with approximately ten clonal replicates per genotype) were planted in the field in 2000 at the INRA's 89 90 experimental unit of Bourran (44°20'N, 0°24'W), located in the south west of France (Saintagne et al., 91 2004). Spacing of trees was 4m x 1.5m at the plantation. In 2012 a systematic thinning was carried out 92 in the parcel, reducing the overall density by one half on average with a total amount of 1130 93 individuals. On average, the annual precipitation was 650.1 mm and the average temperature was 94 13.9°C from 2014 to 2017, i.e. during the four years of monitoring (Table 1).

Genetic linkage maps were established for both parents (3P and A4) using gene-based SNP
(Bodénès et al., 2016). Using the JoinMap procedure, a subset of SNP markers evenly distributed along
the 12 linkage groups (LG) was selected to reconstruct two new parental linkage maps for QTL analysis.
In total, the male and female linkage maps used for QTL detection contain 341 and 345 markers,
respectively.

100 Monitoring reproduction

During spring 2014, when the trees were 16 years old since they were vegetatively propagated from the ortet (and 23 since seed production), we set up nets to collect acorns at 1 m above the ground under the whole canopy of each tree (Figure S1). The nets were stretched at the individual scale every

104 two rows within the design and the trees which were very close to one another within a given row (i.e 105 with no trees cut between them after thinning) were not sampled for recording reproduction, as their 106 canopies were intermingled (Figure S1). In total seed crop was assessed on 173, 331, 339 and 337 107 individuals (clonal replicates) in 2014, 2015, 2016 and 2017 corresponding from 117 to 170 different 108 genotypes over the four years (Table 1). In what follows the full sib genotypes will be called clones. 109 Trees were planted within a randomized incomplete block design comprising 8 blocks with 47 ± 6 110 individuals per block. The number of clonal replicates per clone per block was one and the overall mean 111 number of replicates used for this study varied between 1.48 and 1.99 (Table 1). To minimize predation 112 and facilitate the acorns sorting, trees were harvested twice during mid-October and mid-November 113 by collecting all the material (litter and acorns) deposited in the nets. Acorns were sorted from the rest 114 of the litter in the laboratory and dried at room temperature.

115 Then, the total amount of seed produced N_{tot} , the total mass of seeds produced M_{tot} in g and the mean 116 mass of one seed M_a in g were assessed. Along the last three years of measurement (2015, 2016 and 2017) we also estimated the coefficient of variation of each individual CV_i and the mean coefficient of 117 118 synchrony ri using the coefficient of Pearson (Buonaccorsi et al., 2003). As the number of trees assessed 119 in 2014 was much lower due to technical constraints related to the stretching of the nets, data of 2014 120 were not used in this analysis. In addition, diameter at breast height (D in mm) and height (H in m) of 121 each tree were measured in 2014 and the ratio between reproduction and growth (N_{tot}/D) were 122 calculated for each year in order to normalize the total seed production relative to the size of the tree.

123 2.2. Statistical analysis

We used a univariate linear mixed effect model to assess the genetic parameter of reproductive andgrowth traits for the four years of measurement:

126
$$Y_{ijk} = \mu + b_i + C_j + \varepsilon_{ijk} \quad [1]$$

where Y_{ijk} denotes the observed phenotypic value of clone replicates *k* of clone *j* in block i, μ the overall mean, b_i the fixed effect associated with block *i*, C_j the random effect associated with clone *j* (genetic effect) and ε_{ijk} the residuals. In addition to assess the multi-annual variability we used an additional univariate linear mixed model for each trait:

131
$$Y_{ijkl} = \mu + b_{ij} + t_l + C_j + \varepsilon_{ijkl}$$
 [2]

where Y_{ijkl} is the observed phenotypic value of clone j in block i and year l and t_l the fixed effect 132 associated to year *I*. For both models, the reproductive traits M_{tot}, N_{tot} and the ratio N_{tot}/D were log 133 134 transformed. Best linear unbiased predictions (BLUPs) of random effects were estimated for each trait 135 and within each year from the two linear mixed effect models. Year-Year correlations were estimated 136 for phenotypic and BLUP values of log (N_{tot} + 1), log (M_{tot} +1), log (N_{tot}/D + 1) and M_a using Pearson 137 coefficient, thus leading to values close to phenotypic and genotypic correlations. In addition, the 138 differences between years for the same traits were tested using an analysis of variance (ANOVA). 139 Finally, genetic correlations were also estimated between reproductive and growth traits, using the BLUPs estimated from the multi-annual model for reproductive traits and the BLUPs estimated in 2014 140 for growth traits. 141

Variances of random clone effects were used to estimate the genetic parameters of each trait. Following Scotti-Saintagne et al. (2004) we assume that the environmental effect was absorbed by the variance among replicates (cuttings) of a same genotype (clone). Considering that the cuttings were full sibs, the clonal variance estimated (σ_c^2) was equal to the within full-sib family (σ_w^2), where:

146
$$\sigma_c^2 = \frac{1}{2}V_A + \frac{3}{4}V_D$$
 [3]

Here, V_A is the additive genetic variance and V_D is the dominance variance. To estimate the narrow sense heritability (h^2) of each trait, which represents the proportion of phenotypic variance that can be attributed to V_A , we considered two cases encompassing the likely range of the dominance variance ($V_D = 0$ and $V_D = \frac{1}{2}V_A$), as V_D is generally lower than V_A in forest trees (Cornelius 1994). Thus we estimated h^2 using:

152 (i)
$$h_0^2 = \frac{2\sigma_c^2}{2\sigma_c^2 + \sigma_\epsilon^2}$$
 and (ii) $h_{1/2}^2 = \frac{\frac{8}{7}\sigma_c^2}{\frac{12}{7}\sigma_c^2 + \sigma_\epsilon^2}$ [4]

153 Confidence intervals of narrow sense heritability values were estimated using the method of Visscher 154 and Godard (2014) applied to a single full sib family, assuming that the true values of heritability and

155 phenotypic variance corresponded to the estimated values.

156

157 In addition, we also estimated the repeatability, expressing the proportion of the phenotypic

158 variance of clonal means due to to the clone effect, using:

159
$$R = \frac{\sigma_c^2}{\sigma_c^2 + \frac{\sigma_c^2}{n_0}} \quad [4]$$

160 where (σ_{ε}^2) is the environmental here residual variance and n_0 is the number of cuttings per clone. 161 Finally, we also estimated the evolvability (Hansen et al., 2011) for the two cases previously described 162 for the estimation of h^2 , using:

163 (*i*)
$$e_0 = \frac{2\sigma_C^2}{m^2}$$
 and (*ii*) $e_{1/2} = \frac{\frac{8}{7}\sigma_C^2}{m^2}$ [6]

were m is the mean of the trait. For N_{tot}, M_{tot} and N_{tot}/D, we estimated the evolvabilities of the non-164 165 transformed data since as reported by Hansen et al. (2011), the additive variance of the log-166 transformed data is approximately equal to the evolvability of the non-transformed scale. Evolvability 167 is a mean standardized measure of the additive variation, in comparison to heritability which is a 168 variance standardized measure of additive variation. It was shown earlier (Hansen et la., 2011)) that 169 the latter might respond more to the environmental variance (which is on the denominator of 170 heritability) than to the additive variance (which is on the numerator). To reduce the noise created by 171 the environmental variance on heritability, Hansen et al. (2011) recommended to estimate evolvability 172 as well.

The analyses were performed in R Studio version 1.0.153 (R core team 2014). All the linear mixed effect
models were fitted by the restricted maximum likelihood (REML) method in the lme4 R package (Bates
et al., 2014; Bolker et al., 2009).

177 2.3. QTL detection

178 Quantitative trait locus (QTL) mapping was performed using Haley-Knott regression (Haley and Knott, 179 1992) in R/qtl package (Broman et al., 2003) with 1cM step using the BLUP values. QTLs were selected 180 using a stepwise model selection approach (Manichaikul et al., 2009) based on a significant threshold 181 at 5% error rate made from 1000 permutations. The 95 % confidence interval was calculated for each 182 selected QTL using Bayesian methods (Manichaikul et al., 2006). The effect of each QTL and the 183 percentage of phenotypic variance explained (PEV) were also estimated. In addition, we also used a two QTL model when the results of one QTL model suggested the likely presence of a second QTL on 184 185 the same linkage group, LG₃ (Figure S3). The scantwo function on R/QTL was used in this case (Figure S4). 186

187 3. Results

188 3.1. Phenotypic and genetic variation of reproductive traits

189 Large phenotypic variation was observed for seed production and size over the four years (Figure 1). 190 Mean annual number of acorns per tree (Ntot) varied between 27.8 and 226.6 seeds while Ntot/D varied 191 between 0.54 and 2.34 seeds per mm and M_{tot} between 16.1 and 385.9 gramme. Similar results can 192 be observed when comparing seed production of a given tree over years (Inter-annual coefficient of variation for N_{tot} amounts to 1.05 ± 0.29, for the four years and 1.07 ± 0.35 for the last three years). 193 194 On average the production of seeds was higher in 2014 and 2017 (Figure 1a, b and d) and the mean mass of one seed was higher in 2014 (Figure 1c) with an average mass of 2.5 g. Furthermore, within a 195 196 given year we observed a large inter-individual variability for all reproductive traits (Figure 1). The large differences between marginal and conditional coefficient of determination (R²_m and R²_c) observed for 197 198 most of the annual and multi-annual traits (trait over the four years of monitoring, see model [2]), 199 show that the clone random effect of the linear mixed effect model explains most of the variability 200 predicted by model 1 for one year traits and by model 2 for multi-annual traits (Table 2). Clonal 201 repeatability varied between 0.22 for Ma to 0.70 for Ntot and Ntot /D, suggesting that the between clonal 202 variance was almost as large as the within clonal variance. One can notice that the difference between R²_m and R²_c, estimated from the multi-annual model of the mean mass of one acorn (M_a), is lower than 203 204 for the other traits (R^2_m = 0.05 and R^2_c = 0.1). This discrepancy is most likely due to the large differences 205 of mean acorn mass between years already pointed out. The narrow sense heritabilities (h_0^2 and $h_{1/2}^2$) 206 estimated for each year were moderate and similar over time for every reproductive trait monitored 207 (Table 2). For each year, the highest heritabilities were observed for N_{tot} and N_{tot}/D and the lowest 208 heritabilities were estimated for M_a, in most cases the heritabilities estimated for growth traits (D and 209 H) were lower than for reproductive traits. In comparison, very high evolvabilities were observed for 210 N_{tot} and M_{tot}, ranging from 0.37 to 1.24 but there were more moderate for Ntot/D. The evolvabilities 211 estimated were much lower for Ma (ranging from 0.0008 and 0.04), H, D nevertheless there are still 212 relatively high compared to the other estimated valued in the literature except for H (Hansen et al., 213 2011). Heritability values of the masting related traits (coefficient of variation CV_i and coefficient of 214 synchrony r_i) were of the same magnitude than values of the reproduction related traits (Table 2)

215 3.2. Phenotypic and genetic correlations between traits over time

216 Phenotypic and genotypic correlations between traits over years were positive. For all six pairwise 217 year-year combinations the phenotypic and genotypic correlations were significant for each trait. The 218 coefficients of Pearson (Figure 2 and 3) were, in most cases, higher for the estimated BLUP values than 219 for the phenotypic values with one exception for M_a (2015-2017), for which the phenotypic correlation 220 (r = 0.33) was higher than the estimated genotypic value (r = 0.31). In average, the year-year genotypic 221 and phenotypic correlations were much lower for M_a than for the three other traits. In addition, the 222 highest r values for the genotypic and phenotypic correlations were observed between 2014 and 2017 223 for $\log(N_{tot}+1)$, $\log(M_{tot}+1)$ and $\log(N_{tot}/D + 1)$, while for M_a, the highest phenotypic correlation was 224 observed between 2014 and 2016 and the highest genotypic correlation between 2016 and 2017. The 225 genotypic correlations of reproductive traits with growth traits (H and D) were positive for M_{tot}, N_{tot} 226 and nearer 0 for M_a and N_{tot}/D (Figure S2).

227 3.3. QTL detection (Table 2)

228 Annual and multi-annual models were used to detect the QTLs of reproductive traits. We detected significant QTLs for all traits related to reproduction and growth but not for CV_i neither for r_i. For M_a 229 230 we detected significant QTLs on both maps (Male and Female), 1 QTL on the female linkage group 1 231 (LG₁F) for 2016, 2017 and for the multi-annual model, and 2 QTLs on LG₇M (male linkage group 7) for 232 2016 on LG₁₁M for the multi-annual model (Table 3, Figure S5). The QTLs detected on LG₁F were, for 233 the three cases, located at 26 cM and the percentage of phenotypic explained variance (PEV) was of 234 the same amount over the years: 9.80 % in 2016, 8.08 % in 2017 and 12.81 % over the four years, but 235 the confidence intervals in 2016 and 2017 were much larger. For traits related to seed production (M_{tot}, 236 N_{tot} and N_{tot}/D) we detected significant QTLs at the same location on LG₃F in 2015, 2016, and 2017 and 237 across the three years with the multi-annual model. The positions were very similar through time and 238 between traits. The highest PEVs on this linkage group were observed for N_{tot}, with 10.94% in 2015, 239 15.01% in 2016, 17.51 % in 2017 and 16.47 % for across the four years. In addition, on $LG_{11}M$, we also 240 detected highly significant QTLs for the same traits than previously in 2014, 2016, and 2017 and across 241 all years. The highest PEVs observed were for N_{tot}/D with respectively 15.38%, 17.54%, 12.32% and 242 14.11% in 2014, 2016, and 2017 and across all years. Another QTL was detected on the male map for 243 traits related to seed production: for N_{tot} in 2014 and N_{tot}/D in 2014 on LG₅M. For both cases the position estimated on the linkage group was 65.03 cM and the PEV amounted to 10.11 % and 13.84 % 244 245 respectively. Finally we also detected a significant QTL for N_{tot}/D in 2015 on $LG_{12}F$ but the PEV 246 estimated was relatively low (7.69 %) compared to the other QTLs.

247

248 4. Discussion

Reproduction in forest tree species is highly variable between and within populations (Haymes and Fox, 2012; Kang et al., 2003; Pérez-Ramos et al., 2014). Although few studies have shown and quantified the genetic contribution to the between population variation of reproductive traits (Santosdel-Blanco et al., 2012), our study is one of the first to investigate the genetic variation within a single population. Using a full-sib family of *Quercus robur*, this study highlights a large variability of reproductive traits at the within population level, and underlines the important contribution of genetic effects. Furthermore, we detected for the first time in forest tree species quantitative trait loci (QTLs) associated with seed production and the mean mass of a seed. Despite a large phenotypic variability over years, the genetic contribution to reproduction was highly correlated between years.

258 4.1. Large genetic variation of reproductive traits

259 The repeated assessment of reproduction related traits over four years resulted in estimates 260 of large phenotypic variability for N_{tot}, M_{tot} and N_{tot}/D and to a lesser extent seed size (M_a), as well as 261 for masting related traits (CV_i and r_i) In addition, the estimated heritability and evolvability values 262 suggested significant and substantial genetic contributions to the phenotypic variability. It is tempting 263 to compare our values of heritability and evolvability to other reported values in trees; however the 264 scarce published tree studies reported broad sense heritabilities (H²), which are larger than narrow 265 sense heritabilities. Nevertheless, our results suggest slightly larger genetic (additive) variance of 266 reproductive traits in oaks than in other species. For example Sivacioglu et al. (2009) reported H² values ranging from 0.18 to 0.38 for cone production in *Pinus sylvestris* while H² was found lower, around 267 268 0.15 in *Pinus pinea* (Mutke et al., 2005). These values are similar to the H^2 estimated for female and 269 male inflorescences in pines and spruces (Bilir et al., 2006; Nikkanen and Ruotsalainen, 2000; Sıvacıoglu 270 et al., 2009). For example H² values estimated for female and male flowering were around 0.38, in 271 Picea abies (Nikkanen and Ruotsalainen, 2000) and 0.12 in Pinus sylvestris (Bilir et al., 2006). Compared 272 to other traits commonly assessed in forest trees and especially in oak, narrow sense heritability values 273 (h²) assessed here on reproduction traits are slightly lower than for phenological traits (Baliuckas and 274 Pliura, 2004; Alberto et al., 2011; Firmat et al., 2017), about the same magnitude than wood density 275 and wood anatomical related traits (Nepveu, 1982, 1984; Mather et al., 1993; Savill et al., 1993), and 276 higher than growth traits (see our results, Jensen et al., 1997; Bogdan, et al. 2004,2017; Barzdajn, 277 2008). Therefore, our results suggest that reproduction traits in trees may undergo evolutionary 278 changes if they are targets of selection under ongoing environmental changes.

279 Over the four years of monitoring, seed production was highly variable and synchronized 280 between trees. This phenomenon, also known as masting or mast-seeding, is characteristic of several 281 forest tree species including oak species (Koenig et al., 1996, 1994). The coefficient of variation (CV_i) 282 and the coefficient of synchrony (r_i) estimated for each individual are common measures of the ability 283 to express multi-annual variability of seed production and synchrony among trees, respectively (Buonaccorsi et al., 2003). The moderate heritability estimated for both CV_i ($h^2_0 = 0.49$ and $h^2_{1/2} = 0.30$) 284 285 and r_i ($h_0^2 = 0.40$ and $h_{1/2}^2 = 0.24$) suggests that the variation of both measures, estimated from 2015 286 to 2017, were partly due to genetic effects. While the period of monitoring was relatively short in our 287 study, these results suggest that masting-related traits might be heritable. To our knowledge, no study 288 has attempted so far to study the genetic determinism of masting while it is commonly assumed that 289 it is an adaptive response to the selection pressure by predators (Kelly and Sork, 2002). Our results 290 should however be considered as very preliminary, as long-term longitudinal monitoring would be 291 necessary to estimate the genetic contribution of masting related traits.

292 Despite the large multi-annual variability and the significant sensitivity of reproduction to 293 environmental changes, the genetic contribution was highly correlated over time. Thus, our results 294 showed that most prolific trees were the same during the four years of monitoring, regardless of the 295 overall level of seed crop within each year. For every trait assessed, heritabilities estimated using the 296 multi-annual model was lower than the heritabilities estimated within single year. These differences 297 may be explained by the increase of the residual variation over years due to the changes of biotic and 298 abiotic conditions over time. Evidence for the inter-annual variation of the residual variance is also 299 suggested by the changes of single year heritabilities over years. Similar trends were also observed in 300 other species; for example broad sense heritability (H²) of female and male inflorescence production 301 in Picea abies (Nikkanen and Ruotsalainen, 2000) and cone production in Pinus sylvestris (Kroon et al., 302 2009) changed substantially over years. Despite variation of the overall mean and variance of 303 reproductive traits over years, phenotypic and genetic correlations between years were quite high

suggesting that monitoring of reproduction for genetic evaluation of clones can be limited to a very
 few numbers of years, if masting is not foreseen as an objective of the study.

306 While it is commonly assumed and observed that fitness-traits have low narrow sense 307 heritabilities (Hoffmann et al., 2016; Price and Schluter, 1991), we obtained relatively high values for 308 reproductive traits which are known to be main components of tree fitness. Merilä and Sheldon, (1999) 309 have shown that contrary to what was admitted before, the low heritability of fitness traits was mostly 310 explained by high residual variances including the environmental and dominance variance. Admittedly 311 we assumed in our estimation of narrow sense heritability that dominance (on a broader scale non 312 additive) variance was lower than additive variance in trees based on earlier reported values (White et 313 al., 2007). Our assumption of low V_D may account for the discrepancy between observed values and 314 predicted values based on evolutionary theory of fitness related traits. However, the underestimation 315 of the residual variance would have a lesser impact on evolvability which is a standardization of the 316 additive variance on the mean of the trait. And evolvability values still suggest that there is large 317 additive variation existing in this oak clonal trial. Indeed, with a few exceptions our reported values of 318 evolvability are in the upper half of all reported values of various species in the meta-analysis of Hansen 319 et al. (2011). An alternative interpretation of the moderate values of genetic variation we found for 320 reproductive traits is that four years assessments of reproduction at a still young stage of oak trees 321 might not provide a relevant proxy of fitness. Hence the traits we measured might not have undergo 322 so far sufficient selection pressures to the point to erode the genetic variation of the trait. Finally the 323 evolutionary implications of our results are still questionable, as estimates of heritability in our study 324 were done in an experimental design where micro-environmental variation is minimized. These 325 estimates should be compared in the future with *in situ* estimates, which can now be obtained by 326 retrieving realized genetic relatedness among trees using large numbers of genetic markers (Lesur et 327 al., 2018; Vinkhuysen et al., 2013).

328 4.2. Genetic architecture

Our QTL detection was implemented in a trial prone to sampling biases known as the Beavis effect (Beavis 1998). Typically when the sample size (number of clones in our experiment) is less than 100, then the statistical power to detect QTLs of small effects are low, and their effects are inflated. Because of our limited sample size, our results should be interpreted with caution. We have likely only detected QTLs with major effects, and their effects may also have been overestimated. Thus the refined genetic architecture (number of QTLs and distribution of their effects) needs still further investigations based on the distribution of allelic effects as was done by Hall et al. 2016

336 Despite statistical limitations for exploring the genetic architecture, our results have important 337 biological implications regarding potential genomic regions containing genes that contribute to the 338 variation of reproduction traits. Indeed, for seed production, significant QTLs on LG₃F and LG₁₁M were 339 repeatedly detected over years, explaining from 8.21 to 18.40 and 10.70 to 17.54 percent of the 340 phenotypic variance, respectively. The inferred position on the linkage groups were almost identical 341 between years. Although we found a significant QTL on LG₁₁M for M_a with the multi-annual model, the 342 other QTLs identified were not co-localized with the QTLs identified for the seed production related 343 traits, even with the QTLs detected for M_{tot}. These genomic regions will be targets for association 344 studies in natural populations to reduce their range within the genome and narrow down the search 345 of candidate genes within the genome of Quercus robur that was recently sequenced (Plomion et al., 346 2016, 2018). As very few studies have attempted to dissect the genetic architecture of tree 347 reproduction, it is difficult to compare our results with earlier reported studies. While Ujino-Ihara et 348 al. (2012) undertook detection of QTLs associated with the production of male strobili in Cryptomeria 349 japonica, to our knowledge, seed production and seed size have never been investigated in forest tree 350 species. In fruit trees, numerous studies investigated the genetic determinism of biennial fruit bearing 351 i.e. the irregular fruit production of a tree over consecutive years. Guitton et al. (2011) found that the 352 QTLs associated with biennial bearing co-localized with genomic regions containing genes involved in 353 floral development (floral integration gene, meristem identity gene and gibberellin oxidase gene). These results illustrate how our results could further lead to the identification of candidate genes by refining genomic regions containing the 3 major QTLs that we detected.

356 Finally, it is worthwhile checking whether the QTLs of reproduction related traits may colocalize 357 with QTLs of other important traits particularly growth and phenology that were investigated earlier 358 on the same QTL mapping pedigree (Scotti-Saintagne, 2004; Derory et al. 2010). While, the likely 359 positive correlation between tree size and non-normalized seed production (N_{tot}) may lead to detect 360 similar QTLs for N_{tot} and growth traits, the cross comparison of our results with QTLs of growth resulted 361 in only one notable co-localization. LG_5M for the total tree height in 2001 and for N_{tot} and N_{tot}/D in 362 2014. Due to the low sample size in 2014 and the absence of repetition for the QTL on LG₅M, the co-363 localization of QTLs of seed production and growth on this region is weak. Apart from this region, no 364 co-localization was observed between growth and reproduction. Potential co-localization may witness 365 negative pleiotropic effects that may support the negative mechanistic trade-off between growth and 366 reproduction that has often been reported in the literature (Camarero et al., 2010; Drobyshev et al., 367 2010; Han et al., 2008; Ishihara and Kikuzawa, 2009; Koenig and Knops, 1998; Monks and Kelly, 2006; 368 Silvertown and Dodd, 1999). The lack of co-localization of QTLs in our study may thus be in line with 369 the more recent hypothesis by Knops et al. (2007) that the negative correlation observed might not be 370 causal, or that the trade-off may only be driven by environmental effects. Furthermore, the positive or 371 non-genetic correlation observed between growth and reproductive traits (Figure S2), confirmed the 372 likely absence of a trade-off between both. The higher correlations overserved between Ntot and 373 Ntot/D with growth traits, suggest here the The cross comparison of our results with QTLs of phenology 374 (bud burst) resulted in only one notable co-localization on one linkage group (LG₁₁M), which would 375 support genetic correlation with leaf phenology, seed production and seed size if the gene effects at 376 the co-localized QTLs are strong.

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386 Authors' contributions

T.C., A.K. conceived the idea for this work; T.C. and B.D. assembled the dataset; T.C and C.B. analyzed
the data; T.C., A.K. wrote the manuscript and C.B. and S.D. revised the manuscript.

389 Data Archiving Statement

- 390 <u>http://mapedigree.pierroton.inra.fr/qmap/</u>
- 391

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Table 1: Description of the experimental design for reproductive and growth traits

		Growth			
	2014	2015	2016	2017	2014
Total number of genotypes	117	170	170	170	206
Total number of individuals (clonal replicates)	173	331	339	337	1130
Mean number of clonal replicates per genotype	1.48	1.95	1.99	1.98	5.46
Maximum amount of clonal replicates per genotype	4	6	6	6	10
Yearly averaged temperature (°C)	14.2	14.0	13.6	13.7	14.2
Yearly sum of precipitation (mm)	792.5	511.5	707.5	589.0	792.5

Table 2: Estimation of the marginal and conditional coefficient of determination (R²m and R²c), the mean clone repeatability (R), the narrow sense heritabilities (h²₀ and

h²_{1/2}) and the evolvabilities (e₀ and e_{1/2}) for each trait and each year monitored (from 2014 to 2017)). The interval of confidence of the narrow sense heritabilities were

620 estimated at 95% using the method from Visscher and Goddard (2014).

Trait	Year	n0	R²m	R ² c	R	h²₀	h ² _{1/2}	e ₀	e _{1/2}
N _{tot}	2014	1.48	0.02	0.43	0.52	0.59 [0.23, 0.95]	0.37 [-0.03, 0.77]	1.02	0.58
N_{tot}	2015	1.95	0.07	0.48	0.61	0.62 [0.32, 0.92]	0.39 [0.05, 0.72]	1.24	0.71
N_{tot}	2016	2.00	0.04	0.33	0.46	0.46 [0.13, 0.78]	0.28 [-0.06, 0.62]	0.76	0.43
N_{tot}	2017	1.98	0.02	0.54	0.69	0.69 [0.41, 0.97]	0.44 [0.11, 0.77]	1.03	0.58
N_{tot}	Multi-Annual	2.00	0.13	0.35	0.70	0.40 [0.07, 0.73]	0.38 [0.04, 0.71]	0.89	0.51
M _{tot}	2014	1.48	0.01	0.41	0.50	0.58 [0.21, 0.95]	0.36 [-0.04, 0.76]	1.16	0.66
M _{tot}	2015	1.94	0.07	0.47	0.60	0.60 [0.30, 0.90]	0.38 [0.04, 0.71]	1.09	0.62
M _{tot}	2016	2.00	0.07	0.32	0.43	0.43 [0.10, 0.76]	0.26 [-0.08, 0.60]	0.78	0.44
M _{tot}	2017	1.98	0.04	0.54	0.69	0.69 [0.41, 0.97]	0.44 [0.11, 0.77]	1.16	0.67
M _{tot}	Multi-Annual	2.00	0.13	0.23	0.49	0.22 [-0.11, 0.55]	0.13 [-0.15, 0.41]	0.65	0.37
Ma	2014	1.48	0.01	0.31	0.39	0.46 [0.07, 0.85]	0.28 [-0.12, 0.68]	0.0211	0.0121
Ma	2015	1.88	0.05	0.21	0.27	0.28 [-0.06, 0.62]	0.17 [-0.14, 0.48]	0.0402	0.0230
Ma	2016	1.93	0.07	0.19	0.22	0.22 [-0.11, 0.55]	0.13 [-0.15, 0.41]	0.0210	0.0120
Ma	2017	1.98	0.21	0.42	0.42	0.42 [0.09, 0.75]	0.26 [-0.08, 0.60]	0.0278	0.0159
Ma	Multi-Annual	1.93	0.05	0.1	0.28	0.10 [-0.15, 0.35]	0.06 [-0.12, 0.24]	0.0014	0.0008
N _{tot} /D	2014	1.48	0.02	0.43	0.52	0.59 [0.22, 0.96]	0.37 [-0.03, 0.78]	0.15	0.08
N _{tot} /D	2015	1.95	0.04	0.5	0.64	0.65 [0.36, 0.94]	0.41 [0.07, 0.74]	0.05	0.03
N_{tot}/D	2016	2.00	0.08	0.46	0.58	0.58 [0.28, 0.88]	0.36 [0.02, 0.70]	0.08	0.04
N _{tot} /D	2017	1.98	0.02	0.54	0.70	0.70 [0.42, 0.98]	0.44 [0.11, 0.77]	0.34	0.19
N _{tot} /D	Multi-Annual	2.00	0.16	0.36	0.68	0.38 [0.04, 0.71]	0.23 [-0.10, 0.56]	0.11	0.06
CVi	Multi-Annual	1.93	0.14	0.42	0.48	0.49 [0.17, 0.81]	0.30 [-0.04, 0.64]	0.0593	0.0339
r _i	Multi-Annual	1.98	0.07	0.3	0.4	0.40 [0.07, 0.73]	0.24 [-0.10, 0.58]	0.0332	0.0190
D	2014	5.46	0	0.15	0.48	0.25 [-0.06, 0.56]	0.15 [-0.13, 0.43]	0.017	0.0098
H	2014	5.46	0	0.23	0.62	0.37 [0.06, 0.68]	0.22 [-0.09, 0.53]	7.76e ⁻⁵	4.44e ⁻⁵

no: mean number of clonal replicates per genotype; *Multi-annual*: trait value over the for years; h^2_0 and $h^2_{1/2}$: narrow sense heritabilities assuming dominance variance = 0 623 and dominance variance = ½ of the additive variance; e_0 and $e_{1/2}$: evolvabilities assuming dominance variance = 0 and dominance variance = ½ additive variance

Table 3: Significant QTLs detected for each trait and their related statistics.

Trait	Year	Parent	n	LG	Position	LOD	BCI	PEV	p.value
	2016	Female	168	1	25	3.76	8.85 - 38.00	9.8	<0.001
	2016	Male	168	7	50.08	2.99	36.00 - 58.87	7.86	<0.001
Ma	2017	Female	169	1	26.47	3.11	12.00 - 48.00	8.07	<0.001
	Multi-Annual	Female	170	1	26.00	5.06	20.81-38.00	12.81	<0.001
	Multi-Annual	Male	170	11	34.29	3.24	25.49-52.00	8.39	<0.001
	2014	Male	117	11	54.01	2.97	15.00 - 58.73	11.03	<0.001
	2015	Female	169	3	32.56	3.29	7 – 37.24	8.53	< 0.001
	2016	Female	169	3	33.8	5.86	13.24 – 36.00	14.75	<0.001
N.A	2016	Male	169	11	23.6	6.34	10.72 – 34.29	15.87	< 0.001
IVItot	2017	Female	170	3	28.00	6.78	24.00 - 33.00	16.77	<0.001
	2017	Male	170	11	23.6	5.12	14.00 - 54.01	12.96	<0.001
	Multi-Annual	Female	170	3	31.00	5.85	12.93 - 34.44	14.65	<0.001
	Multi-Annual	Male	170	11	23.6	6.16	14.00 - 34.00	15.36	<0.001
-	2014	Male	117	5	65.03	2.7	45.00 - 72.24	10.11	<0.001
	2014	Male	117	11	54.01	3.02	13 - 58.73	11.24	<0.001
	2015	Female	170	3	32.56	4.28	8.86 - 36.62	10.94	<0.001
	2016	Female	170	3	31	6	13.24 – 35.05	15.01	<0.001
N _{tot}	2016	Male	170	11	23	5.81	10.00 - 33.00	14.56	<0.001
	2017	Female	170	3	29.00	7.76	25.00 - 32.00	17.51	<0.001
	2017	Male	170	11	23.6	4.21	10.00 - 54.01	10.78	<0.001
	Multi-Annual	Female	170	3	31.63	6.64	13.24-34.75	16.47	<0.001
	Multi-Annual	Male	170	11	23.00	4.73	10.00 - 52.00	12.04	<0.001
-	2014	Male	117	5	65.03	3.78	56.78 - 72.24	13.84	<0.001
	2014	Male	117	11	22	4.24	13.00 - 56.00	15.38	<0.001
	2015	Female	170	3	32.6	3.24	4.00 - 39.13	8.41	<0.001
	2015	Female	170	12	19	2.95	10.00 - 30.69	7.69	< 0.001
	2016	Female	170	3	31	5.51	16.00 - 36.62	13.86	<0.001
N _{tot} /D	2016	Male	170	11	10.72	7.12	10.00 - 32.00	17.54	<0.001
	2017	Female	170	3	30.00	7.50	25.19 - 33.00	18.40	<0.001
	2017	Male	170	11	23.6	4.85	10.00 - 54.01	12.32	< 0.001
	Multi-Annual	Female	170	2	30.98	3.23	16.58 - 41.00	8.10	<0.001
	Multi-Annual	Female	170	3	32.00	3.28	17.00 - 40.00	8.21	<0.001
	Multi-Annual	Male	170	11	22.00	5.56	10.72 - 33.00	13.54	< 0.001

- *Year*: year of monitoring; *Multi-annual*: trait value over the four years; *Parent*: Female or male genetic map; *n*: number of clonal replicates per genotype; *LG*: Linkage group;
- *Position*: Position of the QTL on LG in cM; *BCI*: Confidence interval of the position at 95% indicated in cM; *PEV*: Percentage of phenotypic explained variance by a QTL;
- 630 p.value: significance level.

632 Figures:

Figure 1: Boxplots of the 4 phenotypic traits represented for each year. Seed production, total mass
of seed produced (g) and seed production per tree diameter (seed.mm⁻¹) were log transformed. The
mean mass of one seed was assessed in grammes.

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Figure 2: Correlation between years, estimated for the phenotypic values of each trait using the
coefficient of Pearson. Phenotypic values were log transformed for seed production (a), total mass of
seed produced (b) and seed production per tree diameter (d).

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Figure 3: Correlation between years, estimated for the genetic values (BLUPs) of each trait using the

642 coefficient of Pearson. Genetic values of seed production (a), total mass of seed produced (b) and seed

643 production per tree diameter (d) were estimated on the log-transformed phenotypic values.











